

# Package ‘bc3net’

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**Title** Gene Regulatory Network Inference with Bc3net

**Author** Ricardo de Matos Simoes [aut, cre],  
Frank Emmert-Streib [aut]

**Maintainer** Ricardo de Matos Simoes <ricardo\_dematossimoes@dfci.harvard.edu>

**Depends** R (>= 3.0.0), c3net, infotheo, igraph, Matrix, lattice

**Description** Implementation of the BC3NET algorithm for gene regulatory network inference (de Matos Simoes and Frank Emmert-Streib, Bagging Statistical Network Inference from Large-Scale Gene Expression Data, PLoS ONE 7(3): e33624, <[doi:10.1371/journal.pone.0033624](https://doi.org/10.1371/journal.pone.0033624)>).

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## Description

The basic idea of BC3NET is to generate from one dataset  $D_s$ , consisting of  $s$  samples, an ensemble of  $B$  independent bootstrap datasets  $D_k$  by sampling from  $D(s)$  with replacement by using a non-parametric bootstrap (Efron 1993). Then, for each generated data set  $D_k$  in the ensemble, a network  $G^{b_k}$  is inferred by using C3NET (Altay 2010a). From the ensemble of networks  $G^{b_k}$  we construct one weighted network  $G^{b_w}$  which is used to determine the statistical significance of the connection between gene pairs. This results in the final binary, undirected network  $G$ .

A base component of BC3NET is the inference method C3NET introduced in Altay (2010a), which we present in the following in a modified form to obtain a more efficient implementation. Briefly, C3NET consists of three main steps. First, mutual information values among all gene pairs are estimated. Second, an extremal selection strategy is applied allowing each of the  $p$  genes in a given dataset to contribute at most one edge to the inferred network. That means we need to test only  $p$  different hypotheses and not  $p(p-1)/2$ . This potential edge corresponds to the hypothesis test that needs to be conducted for each of the  $p$  genes. Third, a multiple testing procedure is applied to control the type one error. In the above described context, this results in a network  $G^{b_k}$ .

## Details

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bc3net.R c3mtc.R makenull.R mimwrap.R getpval.R mat2igraph.R

## Author(s)

Ricardo de Matos Simoes <r.dematossimoes@qub.ac.uk> Frank Emmert-Streib <f.emmert-streib@qub.ac.uk>  
Maintainer: Ricardo de Matos Simoes <r.dematossimoes@qub.ac.uk>

## References

de Matos Simoes R, Emmert-Streib F., Bagging statistical network inference from large-scale gene expression data., PLoS One. 2012;7(3):e33624. Epub 2012 Mar 30.

## See Also

C3NET, MINET, INFOTHEO

## Examples

```
data(expmat)
bnet=bc3net(expmat)
```

```
data(expmat)
cnet=c3mtc(expmat)
```

---

bc3net

*Bc3net gene regulatory network inference*


---

## Description

The basic idea of BC3NET is to generate from one dataset  $D_s$ , consisting of  $s$  samples, an ensemble of  $B$  independent bootstrap datasets  $D_k$  by sampling from  $D(s)$  with replacement by using a non-parametric bootstrap (Efron 1993). Then, for each generated data set  $D_k$  in the ensemble, a network  $G^b_k$  is inferred by using C3NET (Altay 2010a). From the ensemble of networks  $G^b_k$  we construct one weighted network  $G^b_w$  which is used to determine the statistical significance of the connection between gene pairs. This results in the final binary, undirected network  $G$ .

A base component of BC3NET is the inference method C3NET introduced in Altay (2010a), which we present in the following in a modified form to obtain a more efficient implementation. Briefly, C3NET consists of three main steps. First, mutual information values among all gene pairs are estimated. Second, an extremal selection strategy is applied allowing each of the  $p$  genes in a given dataset to contribute at most one edge to the inferred network. That means we need to test only  $p$  different hypotheses and not  $p(p-1)/2$ . This potential edge corresponds to the hypothesis test that needs to be conducted for each of the  $p$  genes. Third, a multiple testing procedure is applied to control the type one error. In the above described context, this results in a network  $G^b_k$ .

## Usage

```
bc3net(dataset, boot=100, estimator="pearson", disc="equalwidth", mtc1=TRUE,
alpha1=0.05, nullit=NA, null=c(), adj1="bonferroni", mtc2=TRUE,
alpha2=0.05, adj2="bonferroni",
weighted=TRUE, igrph=TRUE, verbose=FALSE)
```

## Arguments

dataset	gene expression dataset where rows define genes and columns samples
boot	default 100 bootstrap datasets are generated to infer an ensemble of c3net gene regulatory networks
estimator	estimators for continuous variables "pearson", "spearman", "kendall", "spearman" estimators for discrete variables "emp", "mm", "sg", "shrink"
disc	required for discrete estimators, method for discretize function (see infotheo package) "equalwidth" (default), "equalfreq", "globalequalwidth"

nullit	<p>nullit defines the size of the generated null distribution vector used for hypothesis testing of significant edges inferred by c3net. The null distribution of mutual information is generated from sample and gene label randomization.</p> <p>number of iterations, where the default is defined by</p> $\text{nullit} = \text{ceiling}(10^5 / (((\text{genes} * \text{genes}) / 2) - \text{genes}))$ <p>genes: number of genes</p>
null	assign alternatively an external null distribution vector
mtc1	consider multiple hypothesis testing for edges inferred by c3net
alpha1	significance level for mtc1
adj1	<p>if mtc1==TRUE default multiple hypothesis testing procedure for c3net inferred edges using "bonferroni" (default)</p> <p>alternatively use "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", "none" (see ?p.adjust())</p>
mtc2	Consider multiple hypothesis testing for edges inferred by bc3net. A binomial test is performed for each gene pair with an ensemble consensus rate >0 consider multiple hypothesis testing for edges inferred by bc3net
alpha2	significance level for mtc2
adj2	Consider multiple hypothesis testing for edges inferred by bc3net. if mtc2==TRUE "bonferroni" is used as multiple hypothesis testing procedure. alternatively use "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", "none"
weighted	A weighted network is returned, where the weights denote the ensemble consensus rate of bc3net.
igraph	A bc3net igraph object is returned.
verbose	Return processing information of running procedures.

## Details

BC3NET Gene regulatory network inference

## Value

'bc3net' returns a gene regulatory network formatted as adjacency matrix, as weighted matrix where the edge weights are defined by the corresponding mutual information values or as undirected weighted or unweighted igraph object.

## Author(s)

de Matos Simoes R, Emmert-Streib F.

## References

- Altay G, Emmert-Streib F. Inferring the conservative causal core of gene regulatory networks. *BMC Syst Biol.* 2010 Sep 28;4:132.
- de Matos Simoes R, Emmert-Streib F. Bagging statistical network inference from large-scale gene expression data. *PLoS One.* 2012;7(3):e33624, Epub 2012 Mar 30, <doi:10.1371/journal.pone.0033624>.

de Matos Simoes R, Emmert-Streib F. Influence of statistical estimators of mutual information and data heterogeneity on the inference of gene regulatory networks. PLoS One. 2011;6(12):e29279. Epub 2011 Dec 29.

### See Also

C3NET c3mtc

### Examples

```
data(expmat)
bnet=bc3net(expmat)
```

---

c3mtc	<i>'c3mtc' gene regulatory network inference using c3net with multiple testing correction procedure</i>
-------	---

---

### Description

We present in the following the inference method C3NET introduced in Altay (2010a) in a modified form to obtain a more efficient implementation. Briefly, C3NET consists of three main steps. First, mutual information values among all gene pairs are estimated. Second, an extremal selection strategy is applied allowing each of the  $p$  genes in a given dataset to contribute at most one edge to the inferred network. That means we need to test only  $p$  different hypotheses and not  $p(p-1)/2$ . This potential edge corresponds to the hypothesis test that needs to be conducted for each of the  $p$  genes. Third, a multiple testing procedure is applied to control the type one error.

In order to determine the statistical significance of the mutual information values between genes we test for each pair of genes the following null hypothesis.

$H_0^I$ : The mutual information between gene  $i$  and  $j$  is zero.

Because we are using a nonparametric test we need to obtain the corresponding null distribution for  $H_0^I$  from a randomization of the data.

The formulated null hypothesis is performed by permuting the sample and gene labels for all genes of the entire expression matrix at once. The vector of the mutual information null distribution is obtained from repeated randomizations for a given number of iterations.

### Usage

```
c3mtc(dataset, null=NULL, mtc=TRUE, adj="bonferroni", alpha=0.05, nullit=NA,
estimator="pearson", disc="none", adjacency=FALSE, igrph=TRUE)
```

**Arguments**

dataset	gene expression dataset where rows define genes and columns samples
nullit	nullit defines the size of the generated null distribution vector used for hypothesis testing of significant edges inferred by c3net. The null distribution of mutual information is generated from sample and gene label randomization. default number of iterations: $\text{nullit} = \text{ceiling}(10^5 / (((\text{genes} * \text{genes}) / 2) - \text{genes}))$ genes: number of genes
estimator	minet package (continuous estimators) "pearson", "spearman", "kendall", "spearman" minet package (discrete estimators) "mi.empirical", "mi.mm", "mi.sg", "mi.shrink" c3net gaussian estimator (pearson) "gaussian" bspline requires installation of "mis_calc" "bspline"
disc	only required for discrete estimators (minet package) "equalfreq", "equalwidth"
mtc	consider multiple hypothesis testing for edges inferred by c3net
adj	if mtc==TRUE default multiple hypothesis testing procedure for c3net inferred edges using "bonferroni" (default) alternatively use "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", "none" (see ?p.adjust())
alpha	significance level for mtc after multiple hypothesis testing correction
adjacency	return an adjacency matrix
igraph	return igraph object
null	If NULL a null distribution vector is generated from a sample label and gene label permutation of the gene expression matrix. For the ensemble inference of one dataset an external null distribution vector is suggested for decreasing running time.

**Value**

'c3mtc' returns a gene regulatory network formatted as adjacency matrix, as weighted matrix where the edge weights are defined by the corresponding mutual information values or as undirected weighted or unweighted igraph object.

**Author(s)**

de Matos Simoes R, Emmert-Streib F.

**References**

- Altay G, Emmert-Streib F. Inferring the conservative causal core of gene regulatory networks. *BMC Syst Biol.* 2010 Sep 28;4:132. PubMed PMID: 20920161; PubMed Central PMCID: PMC2955605.
- de Matos Simoes R, Emmert-Streib F. Bagging statistical network inference from large-scale gene expression data. *PLoS One.* 2012;7(3):e33624. Epub 2012 Mar 30. PubMed PMID: 22479422; PubMed Central PMCID: PMC3316596.
- de Matos Simoes R, Emmert-Streib F. Influence of statistical estimators of mutual information and data heterogeneity on the inference of gene regulatory networks. *PLoS One.* 2011;6(12):e29279. Epub 2011 Dec 29. PubMed PMID: 22242113; PubMed Central PMCID: PMC3248437.

**See Also**[c3](#)**Examples**

```
data(expmat)
net=c3mtc(expmat)
```

enrichment

*Function that performs a functional enrichment analysis based on a one-sided Fisher's exact test (hypergeometric test).*

**Description**

For a given set of candidate genes, reference genes and a list object of gene sets (e.g. Gene Ontology terms or gene sets from pathways) a one-sided Fisher's exact test ("greater") is performed for each gene set in the collection.

**Usage**

```
enrichment(genes, reference, genesets, adj = "fdr", verbose = FALSE)
```

**Arguments**

genes	A character vector of gene identifiers used as a candidate gene list that is assessed by the functional enrichment analysis. The candidate gene list is a subset of the reference gene list.
reference	A character vector of gene identifiers used as the reference gene list. Note all candidate genes are included in the reference gene list.
genesets	A named list object of a collection of gene sets. The identifiers used for the candidate and reference genes need to match the identifier types used for the gene sets. An example list of gene sets is given in <code>data(exgenesets)</code> showing an example list object of gene sets from pathways with gene symbols. <pre>\$'Reactome:REACT_115566:Cell Cycle' [1] "APITD1" "TAOK1" "CDC23" [4] "RB1" "PRKCA" "HIST1H4J" [7] "MCM10" "PPP1CC" "NUP153" ... \$'Reactome:REACT_152:Cell Cycle, Mitotic' [1] "APITD1" "TAOK1" "CDKN2C" [4] "RB1" "PRKCA" "MCM10" [7] "HIST1H2BH" "NUP153" "TUBGCP3" [10] "APEX1" "RPA2" "PRKACA" ...</pre>
adj	The default value is <code>&lt;fdr&gt;</code> (False discovery rate using the Benjamini-Hochberg approach). Multiple testing correction based on the function <code>stats::p.adjust()</code> with available options for "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr" and "none"
verbose	The default value is <code>&lt;FALSE&gt;</code> . If this option is set <code>&lt;TRUE&gt;</code> the number and name of the gene sets during their processing is reported.

## Details

The enrichment analysis is based on a one-sided Fisher's exact test.

## Value

The function returns a data.frame object with the columns

"TermID" the given name of a gene set  $i$  from the named gene set collection list object  $S$ . "genes" the number of candidate genes present in the gene set  $i$  "all" the number of all genes present in the gene set  $i$  "pval" the nominal p-value from a one-sided fisher's exact test "padj" the adjusted p-value to consider for multiple testing

## Author(s)

Ricardo de Matos Simoes

## References

de Matos Simoes R, Tripathi S, Emmert-Streib F. Organizational structure and the periphery of the gene regulatory network in B-cell lymphoma. *BMC Syst Biol.* 2012; 6:38.

Inference and Analysis of Gene Regulatory Networks in R: Applications in Biology, Medicine, and Chemistry, DOI: 10.1002/9783527694365.ch10 In book: *Computational Network Analysis with R*, 2016, pp.289-306

## Examples

```
# In the following candidate genes are defined from a
# giant connected component of an example igraph network
# where we use the remaining genes of a given network
# as a reference list.

data(exanet)
data(exgensets)

candidate=V(getgcc(exanet))$name
reference=V(exanet)$name

# hypergeometric test is performed for
# each defined set of genes in the
# list object exgensets
tab.hypg=enrichment(candidate, reference, exgensets, verbose=TRUE)
```



---

`exanet`*Example gene regulatory network for testing purposes only*

---

**Description**

The network consists of 814 genes and 1,103 interactions.

**Usage**

```
data(exanet)
```

**Format**

An undirected weighted igraph object with 814 vertices and 1,103 edges.

**References**

de Matos Simoes R, Emmert-Streib F, "Bagging statistical network inference from large-scale gene expression data" PLoS One. 2012;7(3):e33624. Epub 2012 Mar 30. PubMed PMID: 22479422; PubMed Central PMCID: PMC3316596.

**Examples**

```
data(expmat)
```

---

`exgensets`*Example gene sets defined from the CPDB database*

---

**Description**

This list data object contains 25 gene sets defined from the CPDB database and is used for testing purposes.

**Usage**

```
data(exgensets)
```

**Format**

A list object with 25 character vectors.

**Details**

An list object of 25 pathway gene sets from CPDB with gene symbols.

**Source**

<http://consensuspathdb.org/>

**References**

A. Kamburov, U. Stelzl, H. Lehrach and R. Herwig, The ConsensusPathDB interaction database: 2013 update, Nucleic Acids Research, Volume 41, Issue D1, Pp. D793-D800

**Examples**

```
data(exgensets)

# The gene set collection from CPDB can be formatted by:
# "CPDB_pathways_genes.tab" is available in the download section in http://consensuspathdb.org/
# cpdb = readLines("CPDB_pathways_genes.tab", warn = FALSE)
# cpdb = lapply(cpdb, function(x) strsplit(x, "\t")[[1]])
# names(cpdb) = sapply(cpdb, function(x) paste(x[3], x[2], x[1], sep=":"))
# cpdb = lapply(cpdb, function(x) x[-c(1:3)])
# cpdb = cpdb[-length(cpdb)]
# cpdb = lapply(cpdb, function(x) strsplit(x, ",")[[1]])
```

---

expmat

*Test gene expression dataset*

---

**Description**

The dataset is test dataset of a gene expression matrix with 100 genes and 100 samples

**Usage**

```
data(expmat)
```

**Format**

A matrix with 100 observations and 100 variables.

**References**

de Matos Simoes R, Emmert-Streib F, "Bagging statistical network inference from large-scale gene expression data" PLoS One. 2012;7(3):e33624. Epub 2012 Mar 30. PubMed PMID: 22479422; PubMed Central PMCID: PMC3316596.

**Examples**

```
data(expmat)
```

---

`getgcc`*Extracting the giant connected component from an igraph object*

---

**Description**

The function uses `igraph::clusters` and `igraph::induced.subgraph` to extract the giant component of an igraph object.

**Usage**

```
getgcc(net)
```

**Arguments**

`net`            An igraph object.

**Details**

In a connected component of an undirected graph all pairs of vertices `u` and `v` are reachable by a path. The giant connected component describes the largest connected component of a graph object.

**Value**

`getgcc` returns an igraph object with the giant connected component of an igraph object.

**References**

G. Csardi and T. Nepusz, The igraph software package for complex network research, InterJournal, Complex Systems,1695, 2006, <http://igraph.org>

**Examples**

```
# The example network "exanet" contains 814 genes with 1103 edges
data(exanet)

# The giant connected component of the example
# network contains 382 genes with 989 edges
getgcc(exanet)
```

gpea

*Gene pair enrichment analysis (GPEA)***Description**

When a network  $G$  contains  $n$  interactions, of which  $k$  interactions are among genes from the given gene set  $S$ , then a  $p$ -value for the enrichment of gene pairs of this gene set  $S$  can be calculated based on a e.g., one-sided Fisher's exact test. For  $p$  genes there is a total of  $N=p(p-1)/2$  different gene pairs (clique graph) with the assumption that all genes within a gene set are associated to each other. If there are  $p_S$  genes for a particular gene set ( $S$ ) then the total number of gene pairs for this gene set is  $m_S=p_S(p_S-1)/2$ .

**Usage**

```
gpea(gnet, genesets, verbose = TRUE, cmax = 1000, cmin = 3,
     adj = "bonferroni")
```

**Arguments**

gnet	igraph object (e.g., inferred from bc3net) of a given network where the gene identifiers $[V(\text{net})\$names]$ correspond to the provided gene identifiers in the reference gene sets.
genesets	A named list object of a collection of gene sets. The identifiers used for the candidate and reference genes need to match the identifier types used for the gene sets. An example list of gene sets is given in <code>data(exgenesets)</code> showing an example list object of gene sets from pathways with gene symbols. <pre>\$'Reactome:REACT_115566:Cell Cycle' [1] "APITD1" "TAOK1" "CDC23" [4] "RB1" "PRKCA" "HIST1H4J" [7] "MCM10" "PPP1CC" "NUP153" ... \$'Reactome:REACT_152:Cell Cycle, Mitotic' [1] "APITD1" "TAOK1" "CDKN2C" [4] "RB1" "PRKCA" "MCM10" [7] "HIST1H2BH" "NUP153" "TUBGCP3" [10] "APEX1" "RPA2" "PRKACA" ...</pre>
verbose	The default value is <code>&lt;FALSE&gt;</code> . If this option is set <code>&lt;TRUE&gt;</code> the number and name of the gene sets during their processing is reported.
cmax	All provided genesets with more than <code>cmax</code> genes will be excluded from the analysis (default <code>cmax=1000</code> ).
cmin	All provided genesets with less than <code>cmin</code> genes will be excluded from the analysis (default <code>cmin&gt;=3</code> ).
adj	The default value is <code>&lt;fdr&gt;</code> (False discovery rate using the Benjamini-Hochberg approach). Multiple testing correction based on the function <code>stats::p.adjust()</code> with available options for "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr" and "none"

**Details**

The enrichment analysis is based on a one-sided Fisher's exact test.

**Value**

The function returns a data.frame object with the columns

"TermID" the given name of a gene set *i* from the named gene set collection list object *S*. "edges" the number of connected gene pairs present a given geneset "genes" the number of candidate genes present in the gene set *i* "all" the number of all genes present in the gene set *i* "pval" the nominal p-value from a one-sided fisher's exact test "padj" the adjusted p-value to consider for multiple testing

**Author(s)**

Ricardo de Matos Simoes

**References**

Inference and Analysis of Gene Regulatory Networks in R: Applications in Biology, Medicine, and Chemistry, DOI: 10.1002/9783527694365.ch10 In book: Computational Network Analysis with R, 2016, pp.289-306

Urothelial cancer gene regulatory networks inferred from large-scale RNAseq, Bead and Oligo gene expression data, BMC Syst Biol. 2015; 9: 21.

**See Also**

See Also as [enrichment](#)

**Examples**

```
data(exanet)
data(exgensets) ## example gene sets from the CPDB database (http://www.consensuspathdb.org)

res = gpea(exanet, exgensets, cmax=1000, cmin=2)
```

---

mimwrap

*Wrapper function for mutual information matrix estimators*

---

**Description**

Mutual information matrix estimation wrapper function for various mutual information estimators. Depends on infotheo package for mutual information estimators on discrete variables.

**Usage**

```
mimwrap(dataset, estimator="pearson", disc="equalwidth", bins = sqrt(ncol(dataset)))
```

**Arguments**

dataset	Data gene expression matrix where rows denote genes (features) and columns samples.
estimator	estimators for continuous variables "pearson" (default), "spearman", "kendall", "spearman" estimators for discrete variables (infotheo package) "emp", "mm", "sg", "shrink"
disc	only required for discrete estimators (see infotheo package) "equalwidth" (default), "globalequalwidth", "equalfreq"
bins	number of bins for the descretize function (infotheo), default $\sqrt{\text{ncol}(\text{dataset})}$

**Details**

A mutual information matrix is estimated from a gene expression data set

**Value**

mimwrap returns a symmetric mutual information matrix for various mutual information estimators.

**References**

Patrick E Meyer, Frederic Lafitte and Gianluca Bontempi, minet: A R/Bioconductor Package for Inferring Large Transcriptional Networks Using Mutual Information, BMC Bioinformatics 2008, 9:461

Carsten O. Daub, Ralf Steuer, Joachim Selbig, and Sebastian Kloska, Estimating mutual information using B-spline functions - an improved similarity measure for analysing gene expression data, BMC Bioinformatics. 2004; 5: 118

de Matos Simoes R, Emmert-Streib F., Bagging statistical network inference from large-scale gene expression data., PLoS One. 2012;7(3):e33624. Epub 2012 Mar 30.

**Examples**

```
data(expmat)
mim <- mimwrap(expmat)
```

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